BACKGROUND

- The London Patient was treated for refractory Hodgkin Lymphoma (stage IVb) with single HSCT with CCR5 Δ 32/ Δ 32 donor cells in 2016¹.
- We reported remission 18 months after analytical treatment interruption using measurements in blood only.
- Here we present data from other tissues and longer follow up.

METHODS

Droplet Digital PCR

We quantified HIV DNA using droplet digital PCR (ddPCR, Bio-Rad) targeting the LTR, gag and integrase region and shown as target copies per million cells tested⁶. RNaseP, a human gene which is present twice in diploid cells, was measured in duplicate to determine the input cell number. In all ddPCR runs, water and donor PBMCs were tested in duplicate as negative controls for the HIV target regions and U1 cells were tested as a positive control. Samples generating a single positive droplet were interpreted as negative based on the occurrence of a sporadic positive droplet in the negative controls (1/40 reactions). We further analyzed samples demonstrating \geq 2 positive droplets in an intact proviral DNA assay (IPDA) essentially as described⁷. The IPDA assay is based on a duplex ddPCR targeting two regions in the viral genome that are present in most intact proviruses: the HIV packaging signal (Ψ) and the Rev Responsive Element (RRE) in Envelope (env).

Ouantitative real time PCR

For each gPCR standards were tested in triplicate, in addition to 2 positive controls in triplicate and 6 negative control wells. 25000 cells worth of DNA (based on an Albumin gPCR) were used per well **HIV-1** antibody responses

Specific HIV-1 antibodies in longitudinal sera samples diluted 1:2 were tested in a qualitative western blot assay (New Lav Blot I, Bio-Rad). Standard and low sensitive (LS) versions of the Vitros anti-HIV-1 assay (Ortho-Clinical Diagnostics) and the limiting avidity antigen assay were measured in same samples

HIV-1, CMV and EBV specific CD4 and CD8 T cell responses

For intracellular cytokine staining (ICS) and peptide stimulation, PBMCs were thawed and resuspended in RPMI complete media. Following, overnight rest at 37°C and 5%CO2, PBMCs were stimulated for 6h with 2µg/ml HIV-1 Gag pools or cytomegalovirus (CMV) pp65 (JPT Peptide Technologies) or the PepTivator® EBV Consensus pool containing 43 peptides of 8-20 aa in length

Mathematical Modelling

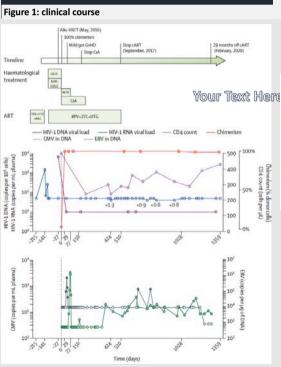
We predicted the probability of rebound using a previouslydeveloped mathematical model and inference approach

RESULTS i

- T cell chimerism maintained at 99%
- pVL below 1 copy/ml to 29 months
- CD4 count 430 at 29 months
- EBV reactivation see figure 1
- CSF viral load <12 copies/ml at 25 months post ATI
- CSF cellular HIV DNA negative at 25 months post ATI
- Semen plasma <12 copies/ml at month 21 post ATI
- Semen cellular HIV DNA negative at 21 months post ATI

References: ¹ Gupta et al, Nature 2019

The London Patient has been in remission for 30 months post ART interruption with evidence of low level HIV-1 DNA 'fossils' in peripheral memory T cells and Lymph node.



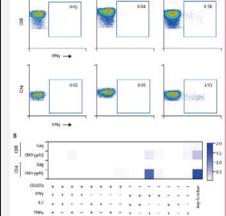


Figure 3. Predictions from mathematical

modelling given no rebound at 29 months

Figure 2. CMV and HIV-1 specific T cell responses

Table 1: ddPCR on tissue compartments

	Sample	Target	Replicates tested	Cells	Copies/million	Clinical
	sample	ranges	(n)	tested (n)	cells	interpretation*
	Lymph node	LTR	14	2068220	33.6	Positive
	Water	LTR	4	0	1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1	
	PBMC	LTR	4	191290	<5.1	
	U1	LTR	4	3091	3409253	
ddPCR						
	Lymph node	Integrase	8	1181840	<0.9	Negative
	Water	Integrase	2	0		
	PBMC	Integrase	2	97240	<10.3	
	U1	Integrase	2	1540	1720000	
	Lymph node	GAG	10	1349700	5.1	Positive
	Water	GAG	2	0		
	PBMC	GAG	2	96177	<10.4	
	Ul	GAG	2	1613	1391197	
IPDA	Lymph node	PSI	14	2068220	1.6	Negative
	Water	PSI	4			
	PBMC	PSI	4	236720	<4.2	
	U1	PSI	4	2860	2289161	
	Lymph node	ENV	14	2068220	26.1	Positive
- 1	Water	ENV	4	0	20.1	PUSITIVE
- 3	PBMC	ENV	4	236720	<4.2	
	U1	ENV	1	236720	2313986	

SPCR = Digital Droplet PCR PDA = intact proviral DNA assa

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ddPCR	Sample	Target	Replicates tested (n)	Cells tested (n)	Copies/million cells	Clinical interpretation*
	lleum	LTR	4	228580	6.7	Negative
	Caecum	LTR	4	358600	2.8	Negative
	Rectum	LTR	4	151800	<6.6	Negative
	Water	LTR	2	0		
	PBMC	LTR	2	94050	<10.6	
	U1	LTR	2	1760	2800000	
	lleum	Integrase	4	201080	<5.0	Negative
	Caecum	Integrase	4	326700	5.4	Negative
	Rectum	Integrase	4	136400	11.3	Negative
	Water	Integrase	2	0		
	PBMC	Integrase	2	97240	<10.3	
	U1	Integrase	2	1540	1720000	

9	Sample	Target	Replicates tested (n)	Cells tested (n)	Copies/million cells	Clinical interpretation*
ddPCR	CD4 cells	LTR	8	424160	<2.4	Negative
	T naive	LTR	8	282920	<3.5	Negative
	T memory	LTR	8	524920	6.7	Positive
	Water	LTR	4	0		
	PBMC	LTR	2	86790	<11.5	
	U1	LTR	2	1562	2887324	
	T memory	PSI	6	286770	21.5	Positive
	Water	PSI	6	0	+	
	PBMC	PSI	2	63690	<15.7	
IPDA	U1	PSI	2	3542	2335404	_
	T memory	ENV	6	886380	6.9	Negative
	Water	ENV	6	0		
	PBMC	ENV	2	63690	<15.7	
	U1	ENV	2	3542	234826	
- 1		One double	e positive droplet in IPD	A (PSI+ENV+)	SHEARING INDEX 0.	24

Negative HIV DNA/RNA results from CSF and semen

Absent HIV specific T cell immune responses

Low level HIV-1 DNA in lymph node and CD4 Tmemory

* = based on \$1 positive droplet per

CONCLUSIONS

 Lymph node gPCR low positive for env and psi Peripheral CD4 memory low positive LTR and env

48 72 96 120 144 168 192 216 240

Time off ART without rebound (months)

80

60

40-

20

24

RESULTS from tissue (table 1)

Gut tissue negative by ddPCR

IPDA negative by ddPCR and qPCR

ALICI

Mathematical modelling (Figure 3): If chimerism is >80% then >90% probability of cure If chimerism is >90% then >99% probability of cure

Lymph node ddPCR low positive for env and LTR



Chimerism (%)

80%

90%

50%



30 month remission post ATI



RESULTS: Immune responses (Figure 2) No CD4 or CD8 responses to HIV whilst CMV responses were observed. Note EBV reactivation in Fig 1

REALTH RESEARCH

Imperial College London

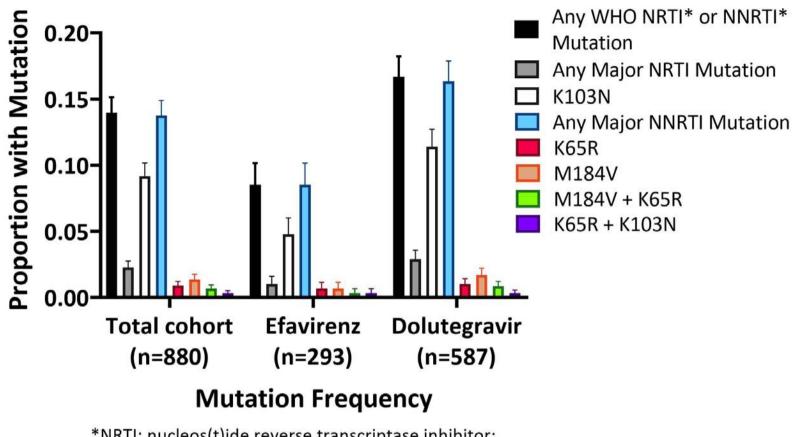
RESULTS (Figure 3)

Table 1. Participant characteristics

Characteristics	Efavirenz Arm (n=293)	Dolutegravir Arms (n=587)	P-value
Female sex, n (%)	169 (58%)	362 (62%)	0.25
Age, median (IQR)	32 (27-37)	32 (27-37)	0.99
Married, n (%)	64 (22%)	114 (19%)	0.41
Tertiary or higher education, n (%)	21 (7%)	56 (10%)	0.23
Employed, n (%)	178 (61%)	365 (63%)	0.55
Baseline CD4, n (%)			
<200 cells/uL	87 (30%)	187 (32%)	
201-350 cells/uL	89 (30%)	176 (30%)	0.60
351-500 cells/uL	62 (21%)	106 (18%)	0.69
>500 cells/uL	55 (19%)	118 (20%)	
Baseline viral load, n (%)			
<10k copies/ml	100 (34%)	186 (32%)	
10k-100k copies/ml	124 (42%)	276 (47%)	0.40
>100k copies/ml	69 (24%)	124 (21%)	
Pre-treatment drug resistance, n (%)*	25 (9%)	98 (17%)	0.001

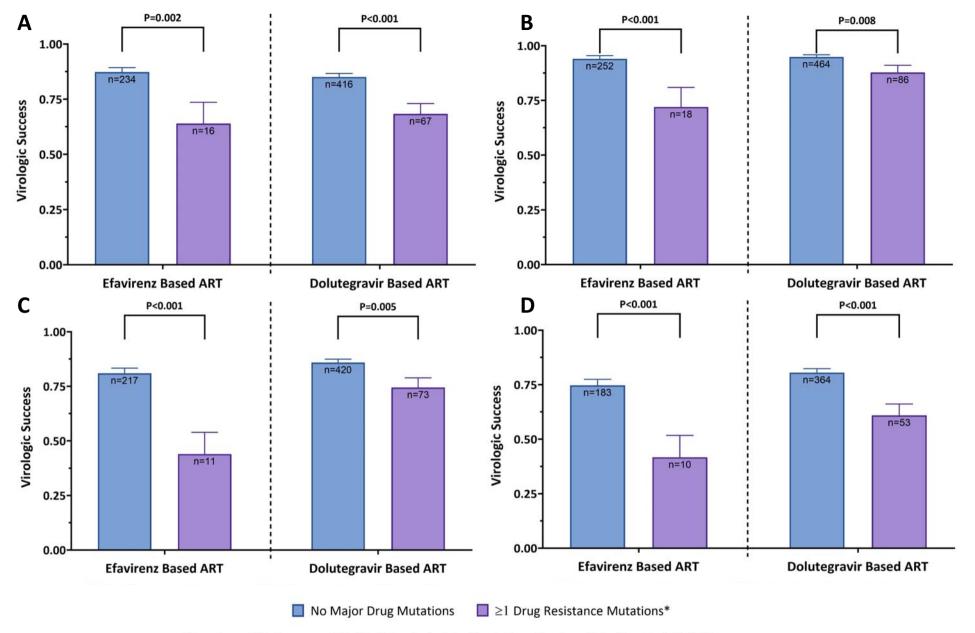
Table 2. Univariable and multivariable regression of predictors of primary outcome							
	Univariable M	odels	Multivariable Model				
	OR (95% CI)	P-value	AOR (95% CI)	<i>P</i> -value			
Female sex	0.85 (0.61- 1.19)	0.34	0.83 (0.55-1.26)	0.38			
Age (per year)	1.05 (1.03-1.07)	< 0.001	1.03 (1.00-1.06)	0.05			
Married	1.61 (1.03-2.51)	0.04	1.09 (0.65-1.83)	0.74			
Tertiary education	1.03 (0.58-1.84)	0.92	0.83 (0.44-1.60)	0.58			
Employed	2.02 (1.46-2.82)	< 0.001	1.62 (1.09-2.42)	0.017			
Baseline CD4							
<200 cells/uL	REF						
201-350 cells/uL	1.20 (0.80-1.81)	0.38	1.12 (0.72-1.96)	0.50			
351-500 cells/uL	1.21 (0.76-1.91)	0.43	0.91 (0.51-1.63)	0.76			
>500 cells/uL	1.20 (0.76-1.90)	0.43	1.01 (0.56-1.84)	0.96			
Baseline viral load							
<10k copies/ml	REF						
10k-100k copies/ml	0.56 (0.38-0.84)	0.005	0.50 (0.30-0.83)	0.008			
>100k copies/ml	0.48 (0.30-0.75)	0.002	0.35 (0.19-0.64)	0.001			
High adherence	0.34 (0.24-0.47)	<0.001	0.34 (0.23-0.50)	<0.001			
Study Arm							
EFV	REF						
DTG	0.71 (0.49-1.01)	0.056	0.88 (0.56-1.39)	0.59			
Arm*WHO PDR			1.82 (0.61-5.42)	0.28			

 Table 2. Univariable and multivariable regression of predictors of primary outcome



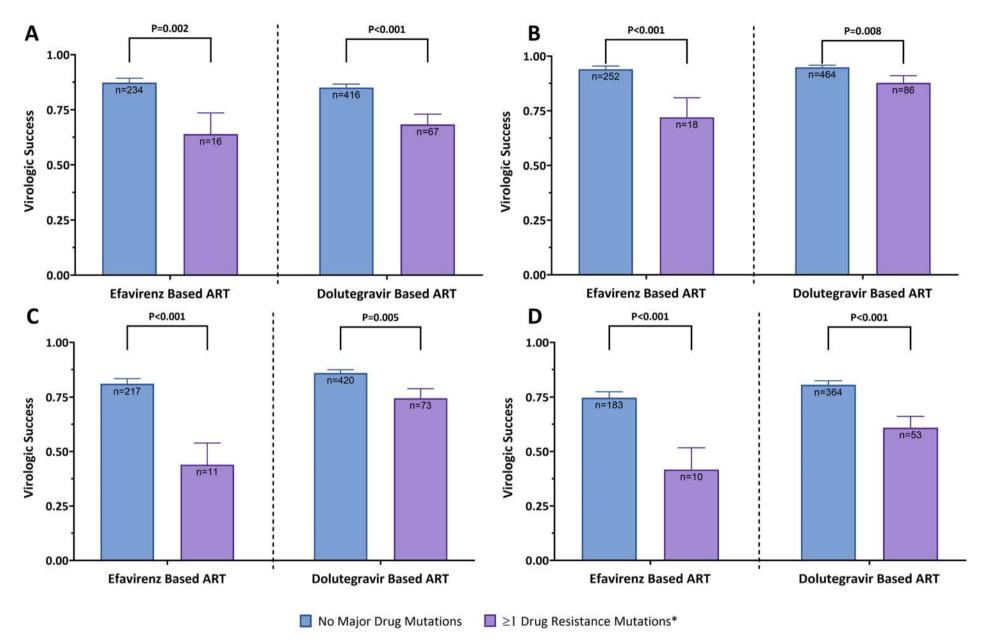
*NRTI: nucleos(t)ide reverse transcriptase inhibitor; NNRTI: non-nucleoside reverse transcriptase inhibitor

Figure 1. Prevalence of pre-treatment resistance in ADVANCE study (n=880)



*Drug resistance defined by presence of World Health Organization-defined Drug Resistance Mutations to Nucleoside or Non-Nucleoside Reverese Transcriptase Inhititors prior to ART Initiation

Figure 2. Primary (A) and Secondary (B) Virologic Outcomes and FDA 48-week (C) and 96-week (D) Snapshot Virologic Suppression by Treatment Regimen and Presence or Absence of WHO Pre-treatment Drug Resistance



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